

# N. Daniel<sup>1</sup>., T. Sivaramakrishnan<sup>2</sup>\*., K. Saravanan<sup>2</sup>., Baby Shalini<sup>2</sup>., Arunjyoti Baruah<sup>2</sup>., Kiruba Sankar, R<sup>2</sup> and S. Dam Roy<sup>21</sup>

ICAR-Central Institute of Fisheries Education, Versova, Andheri West, Mumbai - 400061, Maharashtra <sup>2</sup>ICAR-Central Island Agricultural Research Institute, Post Box No. 181, Port Blair - 744105, Andaman and Nicobar Islands Corresponding Author Email: sivaraman.fish@gmail.com

## Abstract

This review commences by briefly describing the importance of microalgae as a potential feed ingredient in the fish diet. Microalgae are a source of nutrients for fish as well as shrimps. Live microalgae for feeding are being practised in the aquaculture for long ago; but the concerns of using this technique are economics, preservation and storage. It is suggested that instead of using microalgae as live feed, dried microalgae can be used in the feed. It has been noticed that, dried microalgae have a capacity to alleviate those major problems inherent with live microalgae for feed and its suitability to replace the conventional protein ingredients in the fish. Moreover, the production system for freeze dried non-living microalgae is commercially available which ensures the cost efficient dried microalgae will be easily accessible in the market for feed application in animals. The review also highlighted some of the recommendations for future research.

## Introduction

Microalgae are tiny organisms composed of nutrients such as carbohydrates, proteins, lipids, micro-nutrients and other biologically active components. They are further rich in colouring agents such as beta carotene, made it to be used for cultivable fishes like salmon and other ornamental fishes to enhance the colour of fish. In general, fish consume it to attain nutrition and other biologically active substances. Using live microalgae in the feed of fishes are very old technique in the aquaculture. It is being mass cultured and used as a live feed for many years. Despite, live microalgae are important to fish, they are not a good choice to the farmers due to its difficulties in manufacturing, drying and storing them (Borowitzka, 1997; Becker, 2004). Due to these disadvantages, live microalgae are criticized and so make farmers reliant on the ready-made usage of commercial feed for the fishes. It is suggested that dried microalgae in the feed can potentially lessen the usage of other demanded ingredients for aquaculture.

Management of aquaculture practices is dependence on cost effective, better-quality ingredients and feeds. Recent years, fish meal and the other main ingredients used in aquaculture are limiting; thus, feed formulators are looking for substitute ingredients. Newly, the costless, improved qualities of dried microalgae are commercially accessible in the market which will entice feed formulators to be incorporated this in the artificial feed (Muller, 2000; Borowitzka, 1997). Current research is relying on the biotechnological approaches to further improving the quality and cost effective freeze-dried microalgae production for its sustainable usage in the fish feeds.

## Commonly used microalgae

Despite, thousands of microalgae exist on the earth; only few strains are used in the aquaculture. This is due to differences in the culturing techniques, toxicity, nutritional importance, shape, size and digestibility (Raja et al., 2004b; Patilet al., 2007). The species such as Isochrysis, Tetraselmis, Chlorella, Pavlova, Chaetoceros, Phaeodactylum, Skeletonema, Thalassiosira and Nannochloropsis are frequently cultured for animal feed. According to Spolaore et al., (2006) most of the commercially cultivable marine fishes retrieve nutrition principally from the microalgae such as Nannochloropsis sp., Pavlova sp. and Isochrysis sp. It is commonly practiced by using the mixture of different microalgal strains for feeding by saying that they are more nutritionally balanced than single strain; thus, resulting in further improvement in the quality and growth performance of the animal.



### Nutritional profile of microalgae

It is the fact that microalgae possess adequate amounts of biologically active nutrients (Pulz& Gross, 2004; Becker, 2004, Gouveia *et al.*, 2008). They are nutritionally enriched with micronutrients and carotenoids as well. They are also rich in high-quality proteins, amino acids and fatty acids such as docosahexaenoic (DHA), eicosapentaenoic (EPA) and arachidonic acid (AA) which are prerequisite to the animal health (Nell *et al.*, 1991). Omega-3 fatty acids and amino acids are the major demand for animal feed as well as pharmacological industries. A key advantage of focusing on microalgae as an ingredient is due to their higher nutrient profile; including omega-3 fatty acids and the presence of essential amino acids. The table shows the nutritional profile of some selected microalgae as compared with high quality fishmeal.

	Nannochloropsis	Isochrysis	Phaeodactylum	High quality		
	oceania	galbana	tricornutum	fishmeal		
Crude protein, %	47.7	20.1	49.0	74.7		
Crude fat %	8.4	16.2	7.4	9.7		
EPA, C20:5	2.3	0.08	2.8	1.5-2.0		
DHA, C22:6	-	1.6	0.02	0.7-1.3		
Amino acid profile (g/16 g N)						
Lysine	4.8	3.1	4.2	6.8		
Methionine	1.8	2.5	2.0	2.5		
Tryptophan	1.7	2.5	1.3	0.7		
Threonine	3.6	4.6	3.7	3.5		
Valine	4.6	6.1	4.6	4.0		
Isoleucine	3.5	5.1	3.8	3.7		
Leucine	6.7	9.2	6.2	6.2		
Phenylalanine	3.9	5.7	4.2	3.3		
Arginine	4.9	4.1	4.4	5.4		

<b>Comparative nutritional</b>	nrofile of microalgae vs	high anglit	v fishmeal (	%)
	prome or microalgae vs.	mgn quant	y nonnear (	201

Source: Overland (2012), Becker (2004), Raja et al., (2004).

## Scope of microalgae in fish diet

Around 30% of the algae production is utilized in the animal feed. The nutritional and toxicological evaluations have proved the suitability of algal biomass as a feed supplement (Becker *et al.*, 2004). According to Spolaore *et al.*, (2006) consumption of microalgae is linked to supplementation of nutrients, antioxidants and polyunsaturated fatty acids (PUFA). Microalgae are naturally adequate with omega 3 fatty acids. During early larval stages, animals need more omega-3 fatty acids. Many studies showed the essentiality of microalgae in the fish hatcheries as the small fish larvae are efficiently utilizing this as a feed. Apart from the role in nutrition, it also acts as a colouring agent to improve the coloration in the fishes. Commonly used feed ingredients is lacking with the carotenoid pigments such as astaxanthin. Hence, in salmon and other ornamental fish feeds its inclusion will be particularly important.

# Study attempts of microalgae as fish feed ingredient

Some efforts have been made by shifting animals from live microalgae to dry microalgae. Coutteau *et al.*, (1993) suggested that the products of marine algae in the feed can potentially replace requirements of protein sources. It has been proved that aquaculture and livestock diets containing dried microalgae as an ingredient enhanced the immunity (Turner *et al.*, 2002), antibacterial and antiviral effect (Michiels *et al.*, 2012), lipid metabolism (Nakagawa, 1997; Guroy *et al.*, 2011), stress mitigation (Nath *et al.* 2012; Sheikhzadeh *et al.*, 2012). A report from the Kyle *et al.*, (1991) indicating that animals fed with microalgae in the diet clearly improved the weight, immunity, fatty acids, vitamins and mineral profiles. Also, a report from the Azaza *et al.*, (2008) showing that tilapia feed formulated with microalgae as ingredient augments the protein efficiency and growth performances. Besides these, Enright *et al.*, (1986) noticed increased growth rates in the shrimp when shrimp feed were supplemented with *Hypnea cervicornis* and *Cryptonemia crenulata*. Feed prepared with Arthrospira which contains 5% to 20% of carotene pigments improves the colouration in the carp. It wasalso established that the supplementation of Spirulina meal @ 0.5% in shrimp feed act as a potent feeding attractant (Montaini *et al.*, 1995).

#### Commercial production systems

Microalgae can be manufactured almost in a wide variety of environments, even the place where the land and/or water is nil (Raja, 2009). It is produced worldwide for a variety of purposes. Production of algal products in the world is 5 million kg/year. Among that 1 million kg is used for aquaculture purposes (Muller-Feuga, 2004). In the past, culture of microalgae had discrepancy in the cost of production, guality, preservation technique and storage which significantly prevents this for aquaculture applications. However, the substantial advancements have been made over the last five decades for the commercial production systems of culturing microalgae. Continuing research discovers the sophisticated tools such as largescale biocoil type tubular photobioreactor for the largescale production of the greater yield of microalgae, which ensures their reliable, sustainable usage to aquaculture. The development also made in order to concentrate, dry and preserve the algal biomass. In addition to the events raised in the photobioreactors for large scale production, post harvesting and processing tools like spray-dryer and algal biomass concentration were developed for the development of freeze-dried microalgae which have more shelf life which will make microalgae as one among the major fish feed ingredient. So, in the future, it would be expected that microalgae can be a potential feed ingredient in the aquaculture.



Microalgae can meet all criteria to be a part of the major fish feed ingredient. However, its supplementation in the artificial diet is criticized as they require larger proportions of biomass. But, now biotechnological tools help in cultivation of higher quantity of algal biomass which attracted feed industry in some areas. Few studies have already proved its capacity to utilize as one of the major feed ingredient in the aquaculture. Therefore, recommendation has to be done to place this as one of the essential feed ingredient in fish diet. If the brief information available from this article would met, it would be expected that microalgae will be more popularized in the fish feed industry as one of the essential feed ingredient.

As the quality differs for different strains, improved variety must be further explored. Future investigation should also be focused on the development of modified variety of strains and improvement in the manufacturing systems. The transgenic variety of microalgae may have noteworthy potential which needs to be studied. Microalgae are naturally high in polyunsaturated fatty acid (e.g. docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA). However, manipulating the processing methods can further improve its content which needs to be thoroughly explored. It is also recommended that further research has to be carried out to utilize microalgae for fish feed application for the sustainability of aquaculture.

### References

- Azaza, M. S., Mensi, F., Ksouri, J., Dhraief, M. N., Brini, B., Abdelmouleh, A., & Kraiem, M. M. (2008). Growth of Nile tilapia (*Oreochromis niloticus L.*) fed with diets containing graded levels of green algae ulva meal (Ulvarigida) reared in geothermal waters of southern Tunisia. *Journal of Applied Ichthyology*, 24(2): 202-207.
- Becker, W (2004). Microalgae for Aquaculture: The Nutritional Value of Microalgae for Aquaculture. In: Richmond, A. (eds.): Handbook of Microalgal Culture: Biotechnology and Applied Phycology. Oxford, pp. 380–391.



- Becker, W. (2004). Microalgae in Human and Animal Nutrition. Handbook of microalgal culture: biotechnology and applied phycology, 312.
- Borowitzka, M. A. (1997). Microalgae for aquaculture: opportunities and constraints. *Journal of Applied Phycology*, **9**(5): 393-401.
- Coutteau, P., Dravers, M., Dravers, P., Leger, P., & Sorgeloos, P. (1993). Manipulated yeast diets and dried algae as a partial substitute for live algae in the juvenile rearing of the Manila clam Tapes philippinarum and the Pacific oyster *Crassostrea gigas*. Production, Environment and Quality. Bordeaux Aquaculture, 92.
- Enright, C. T., Newkirk, G. F., Craigie, J. S., & Castell, J. D. (1986). Evaluation of phytoplankton as diets for juvenile *Ostrea edulis L. Journal of Experimental Marine Biology and Ecology*, **96**(1): 1-13.
- Guroy, D., Guroy, B., Merrifield, D. L., Ergun, S., Tekinay, A. A., & Yigit, M. (2011). Effect of dietary Ulva and Spirulina on weight loss and body composition of rainbow trout, *Oncorhynchus mykiss* (Walbaum), during a starvation period. *Journal of Animal Physiology and Animal Nutrition*, **95**(3): 320-327.
- Kyle, D. J., & Gladue, R. (1991). Eicosapentaenoic acids and methods for their production. World Patent, vol. 9, pp. 114-427.
- Michiels, J., Skrivanova, E., Missotten, J., Ovyn, A., Mrazek, J., De Smet, S., & Dierick, N. (2012). Intact brown seaweed (*Ascophyllum nodosum*) in diets of weaned piglets: effects on performance, gut bacteria and morphology and plasma oxidative status. *Journal* of Animal Physiology and Animal Nutrition, **96**(6): 1101-1111.
- Montaini, E., Zittelli, G. C., Tredici, M. R., Grima, E. M., Sevilla, J. F., & Perez, J. S. (1995). Long-term preservation of *Tetraselmis suecica*: influence of storage on viability and fatty acid profile. *Aquaculture*, **134** (1): 81-90.

- Muller-Feuga, A. (2000). The role of microalgae in aquaculture: situation and trends. *Journal of Applied Phycology*, **12** (3-5): 527-534.
- Muller-Fuega, A. (2004). Microalgae for aquaculture: the current global situation and future trends. Handbook of microalgal mass cultures. CRC Press Inc. Boca Raton, Florida, 352-364.
- Nakagawa, H. (1997). Effect of dietary algae on improvement of lipid metabolism in fish. *Biomedicine* & pharmacotherapy, **51**(8): 345-348.
- Nath, P. R., Khozin-Goldberg, I., Cohen, Z., Boussiba, S., &Zilberg, D. (2012). Dietary supplementation with the microalgae Parietochlorisincisa increases survival and stress resistance in guppy (*Poecilia reticulata*) fry. *Aquaculture Nutrition*, **18** (2): 167-180.
- Nell, J. A., & Connor, W. A. (1991). The evaluation of fresh algae and stored algal concentrates as a food source for Sydney rock oyster, *Saccostrea commercialis* (Iredale & Roughley), Iarvae. *Aquaculture*, **99** (3): 277-284.

Overland (2012). APC. Unpublished.

- Patil, V., Kallqvist, T., Olsen, E., Vogt, G., & Gislerod, H. R. (2007). Fatty acid composition of 12 microalgae for possible use in aquaculture feed. *Aquaculture International*, **15**(1): 1-9.
- Pulz, O., & Gross, W. (2004).Valuable products from biotechnology of microalgae. *Applied Microbiology* and Biotechnology, **65**(6): 635-648.
- Raja, R. (2009). Microalgae [*Pourriel probable*] a column in the IInd Chapter in 'Un monde invisible' edited by Laurence Bordenave, Publisher: Aubanel-La Martinie`re, La Martinie`re Group, ISBN: 978-2-7006-0670-6, France, pp 124–126 (French).
- Raja, R., Anbazhagan, C., Lakshmi, D., & Rengasamy,
  R. (2004). Nutritional studies on *Dunaliella* salina (Volvocales, Chlorophyta) under laboratory conditions. *Seaweed Res Utili*, 26: 127-146.

Daniel et al.



- Sheikhzadeh, N., Tayefi-Nasrabadi, H., Oushani, A. K., & Enferadi, M. H. N. (2012). Effects of *Haematococcus pluvialis* supplementation on antioxidant system and metabolism in rainbow trout (*Oncorhynchus mykiss*). *Fish Physiology and Biochemistry*, **38** (2): 413-419.
- Sousa, I., Gouveia, L., Batista, A. P., Raymundo, A., & Bandarra, N. M. (2008). Microalgae in novel food products. *Food Chemistry Research Developments*, 75-112.
- Spolaore, P., Joannis-Cassan, C., Duran, E., & Isambert, A. (2006).Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*, **101** (2): 87-96.
- Turner, J. L., Dritz, S. S., Higgins, J. J., & Minton, J. E. (2002). Effects of *Ascophyllum nodosum* extract on growth performance and immune function of young pigs challenged with *Salmonella typhimurium*. *Journal of Animal Science*, **80** (7): 1947-1953.